A NEW XANTHONE C-GLUCOSIDE, POSITION ISOMER OF MANGIFERIN, FROM

ANEMARRHENA ASPHODELOIDES BUNGE

Masakazu Aritomi

Faculty of Education, Kumamoto University

Kurokami-machi, Kumamoto

and

Toshio Kawasaki

Faculty of Pharmaceutical Sciences, Kyushu University

Katakasu, Fukuoka

(Received in Japan 24 January 1969; received in UK for publication 6 February 1969)

Mangiferin (I) which has recently been assigned the conclusive structure 1, 3, 6, 7-tetrahydroxyxanthone 2-C- β -D-glucopyranoside (1) was first isolated from <u>Mangifera indica</u> L. (Anacardiaceae), later from several different families of plants (2) and the occurrence in the rhizomes and the aerial parts of <u>Anemarrhena asphodeloides</u> Bunge (Liliaceae) was reported by Morita <u>et al</u> (3). However, except mangiferin and its 3-methylether (4), no xanthone C-glycoside has been reported, while a number of flavonoid C-glucosides appear in the literatures and among them there are some pairs of coexisting position isomers, such as orientin (II) and isoorientin (III)(5).

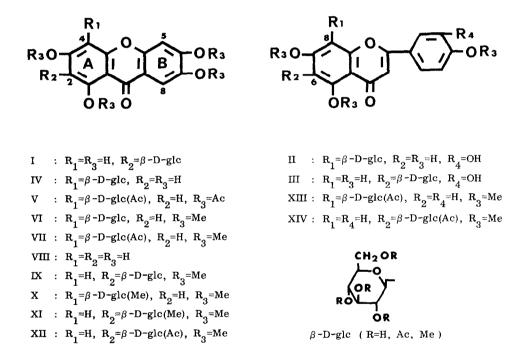
This paper deals with a new xanthone C-glucoside, named isomangiferin (IV), which accompanies I in the aerial parts of <u>A.asphodeloides</u> and is defined as 1,3,6,7-tetrahydroxyxanthone $4-C-\beta-D-glucopyrano$ side, a position isomer of I.

The methanolic extractives of the fresh materials collected in May were subjected to repeated fractional recrystallization from MeOH and 50% (v/v) aq dioxane to give IV, decomposed over 260° without melting, [α]_D + 5.5° (pyridine), Rf 0.50, C₁₉H₁₈O₁₁^{-1/2} H₂O,^{*} as a minor constituent along with I, decomposed

^{*} Rf values were taken on PPC (solvent, 30% (v/v) AcOH). Analytical data were in good agreement with the molecular formulae indicated. NMR spectra were determined at 60 Mcps in CDCl₃ solution with TMS as an internal reference and chemical shifts (δ) are given in ppm.

over 260° without melting, $[\alpha]_{D} + 37.6^{\circ}$ (pyridine), Rf 0.60.

IV showed the UV spectrum and color reactions with Mg-HCl and FeCl₃ almost identical with those of 1, and afforded octaacetate (V), $C_{35}H_{34}O_{19}$, m.p. 203-204° (from CHCl₃-hexane), $[\alpha]_D$ -64.7° (CHCl₃), tetramethylether (VI), $C_{23}H_{26}O_{11}$, m.p. 292-293° (dec) (from aq MeOH), $[\alpha]_D$ -13.2° (pyridine) and tetramethylether tetraacetate (VII), $C_{31}H_{34}O_{15}$, m.p. 142-143° (from CHCl₃-hexane), $[\alpha]_D$ + 9.8° (CHCl₃).



IV resisted an usual acid hydrolysis, but refluxing with HI gave 1, 3, 6, 7-tetrahydroxyxanthone (VIII) and oxidation with FeCl_3 afforded arabinose and glucose (PPC). The NMR spectrum ^{*} of V exhibited three aromatic protons and four acetoxyl groups each on xanthone and sugar moieties (6,7). Oxidation of IV, I and 1-phenyl-1-deoxy- β -D-glucopyranose with KMnO₄ in alkaline medium (8) provided equally D-arabonic acid γ -lactone, m.p. 92-93° ($H_2SO_4^-$ bath) or 133-134° (Kofler) (from EtOAc-benzene), $[\alpha]_D^+$ 69.7° (H_2O) (9) and VI consumed, in common with I tetramethylether (IX), two moles of periodate without vielding formaldehyde.

Therefore IV is regarded as one of the C-D-glucopyranosides of VIII.

IV permethylether (X), m.p. 167° (from aq MeOH), prepared by exhaustive methylation of VI according to the Kuhn method (10) showed in its NMR spectrum (measured at 60°)^{**} eight methoxyl groups, one α -anomeric proton of the sugar moiety (δ 4.97, d, J=10 cps) and three aromatic protons as sharp singlets (δ 7.65, 6.83 and 6.43), indicating that the D-glucopyranosyl residue is β -linked with a carbon atom of the A ring of VIII. The chemical shifts of the two aromatic protons (δ 7.65 and 6.83) of X were in good agreement with those of the C₈-H (δ 7.61) and C₅-H (δ 6.80) of I permethylether (XI), while that of the third one (δ 6.43) appeared at a significantly higher field than that of the C₄-H (δ 6.64) of XI. Since it is known that the C₂-H in many derivatives of VIII is observed at the higher field than that of C₄-H (δ 6.7), the signal at δ 6.43 is assigned to the C₂-H and, hence, the C-glucosidic linkage in X is considered to be located at C-4. The fact that IV was negative and I was positive to the Gibbs reagent (11) supports the assignment.

Consequently IV is formulated as 1, 3, 6, 7-tetrahydroxyxanthone $4-C-\beta$ -D-glucopyranoside.

Recently Prox (12) has reported the mass spectral differences between flavonoid 6-C- and 8-C-glucosides in the free state. Although the spectra of IV and I failed to give any particular information, VII showed in the mass spectrum the molecular ion (m/e 646) as the base peak and the $[M-59]^+$ ion with a relative intensity of 12%, and I tetramethylether tetraacetate (XII) gave the $[M-59]^+$ ion as the base peak and the molecular ion as a minor one (relative intensity, 10%). These data were in good accordance respectively with those of vitexin trimethylether tetraacetate (XIII) (M⁺ 100%, $[M-59]^+$ 9%) and of the corresponding derivative (XIV) of isovitexin (M⁺ 5%, $[M-59]^+$ 100%). Furthermore the empirical rules that the flavonoid 6-C-glucosides show the higher Rf values on PPC (13) and the larger optical rotations (14) than those of the isomeric 8-C-glucosides hold true in I and IV.

It is noted that the relation between xanthone 2-C- and 4-C-glucosides corresponds to that between flavonoid 6-C- and 8-C-glucosides.

** The NMR spectrum of X recorded at 22° showed the anomeric proton as a broad peak at about 5.0.

REFERENCES

- V.K. Bhatia and T.R. Seshadri, <u>Tetrahedron Letters</u>, 1741 (1968); P. E. Nott and J. C. Roberts, <u>Phytochemistry</u>, <u>6</u>, 1597 (1967), and the references cited therein.
- (2) R.A. Finnegan, R.A. Stephani, G. Ganguli, S. N. Ganguly and A.K. Bhattacharya, <u>J. Pharm. Sci.</u>, <u>57</u>, 1039 (1968), and the references cited therein.
- (3) N. Morita, M. Shimizu and M. Fukuta, <u>Yakugaku Zasshi</u>, <u>85</u>, 374 (1965).
- (4) M. Aritomi and T. Kawasaki, Chem. Pharm. Bull. (Tokyo), 16, 760 (1968).
- (5) B.H.Koeppen, C.J.B.Smit and D.G.Roux, Biochem.J., 83, 507 (1962); M.Aritomi, <u>Yakugaku</u>
 <u>Zasshi</u>, <u>83</u>, 737 (1963); M.Yasue, M.Itaya, H.Oshima and S.Funahashi, <u>ibid.</u>, <u>85</u>, 553 (1965);
 H.Wagner, J.Patel and L.Hörbammer, <u>Zeitschrift für Naturforsch.</u>, 22b, 988 (1967).
- (6) D. Billet, J. Massicot, C. Mercier, D. Anker, A. Matschenko, C. Mentzer, M. Chaigneau, G. Valdener and H. Pacheco, <u>Bull, Soc. Chim. France</u>, 3006 (1965).
- (7) L.J. Haynes and D.R. Taylor, <u>J. Chem. Soc</u>. (C), 1685 (1966).
- (8) K. Funaoka, Abstracts of Papers, the 64th Meeting of Japanese Forestry Soc. (Tokyo), 337 (1955).
- (9) F.W. Jensen and F.W. Upson, J.Am. Chem. Soc., 47, 3019 (1925).
- (10) R. Kuhn, H. Trischmann and I. Löw, Angew. Chem., 67, 32 (1955).
- (11) H.D.Gibbs, J.Biol. Chem., 72, 649 (1927).
- (12) A. Prox, <u>Tetrahedron</u>, <u>24</u>, 3697 (1968).
- (13) E.C. Bate-Smith and T.Swain, Chem. Ind., 1132 (1960).
- (14) W. E. Hillis and D. H. S. Horn, Aust, J. Chem., 18, 531 (1965).